



# Standard Test Method for Determining the Bacteria-Eliminating Effectiveness of Healthcare Personnel Hand Rub Formulations Using Hands of Adults<sup>1</sup>

This standard is issued under the fixed designation E2755; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon ( $\epsilon$ ) indicates an editorial change since the last revision or reapproval.

## 1. Scope

1.1 This test method is designed to determine the activity of healthcare personnel hand rubs, (also known as hand rubs, hygienic hand rubs, hand sanitizers, or hand antiseptics) against transient microbial skin flora on the hands after a single application and after repeated applications.

1.2 Performance of this procedure requires the knowledge of regulations pertaining to the protection of human subjects (see 21 CFR Parts 50 and 56).

1.3 This test method should be performed by persons with training in microbiology, in facilities designed and equipped for work with potentially infectious agents at biosafety level 2.<sup>2</sup>

1.4 *Units*—The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.

1.5 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.* For more specific precautionary statements, see 8.2.

## 2. Referenced Documents

### 2.1 ASTM Standards:<sup>3</sup>

**E1054 Test Methods for Evaluation of Inactivators of Antimicrobial Agents**

<sup>1</sup> This test method is under the jurisdiction of ASTM Committee E35 on Pesticides, Antimicrobials, and Alternative Control Agents and is the direct responsibility of Subcommittee E35.15 on Antimicrobial Agents.

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<sup>2</sup> CDC-NIH, *Biosafety in Microbiological and Biomedical Laboratories*, 5th ed., U.S. Department of Health and Human Services, U.S. Government Printing Office, Washington, DC, 2007.

<sup>3</sup> For referenced ASTM standards, visit the ASTM website, [www.astm.org](http://www.astm.org), or contact ASTM Customer Service at [service@astm.org](mailto:service@astm.org). For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

**E1174 Test Method for Evaluation of the Effectiveness of Health Care Personnel Handwash Formulations**

**E2276 Test Method for Determining the Bacteria-Eliminating Effectiveness of Hygienic Handwash and Handrub Agents Using the Fingertips of Adults**

**E2756 Terminology Relating to Antimicrobial and Antiviral Agents**

### 2.2 Other Standards:

**AATCC Test Method 147 2004 Antibacterial Activity Assessment of Textile Materials: Parallel Streak Method<sup>4</sup>**

**21 CFR Parts 50 and 56 Protection of Human Subjects; Institutional Review Boards<sup>5</sup>**

## 3. Terminology

3.1 *Definitions*: For definitions of terms used in this document, see Terminology E2756.

### 3.2 Definitions of Terms Specific to This Standard:

3.2.1 *healthcare personnel handrub, n*—an antimicrobial gel, foam, liquid, spray, or wipe, applied by rubbing to reduce the transient microbial skin flora on hands that are not visibly soiled, and which does not require a post-treatment water rinse. Such agents may also be referred to as hand rubs, hygienic hand rubs, or hand antiseptics.

3.2.2 *healthcare personnel handwash, n*—a cleanser or waterless agent intended to reduce transient microbial skin flora on the hands.

3.2.3 *test bacteria, n*—an applied inoculum of bacteria that has characteristics which allow it to be readily identified. Test bacteria are used to simulate a topical transient microbial contaminant. This may also be referred to as a test organism, marker organism, simulant, or contaminant.

3.2.4 *test material, n*—a product or formulation which incorporates an antimicrobial ingredient(s).

<sup>4</sup> Available from American Association of Textile Chemists and Colorists (AATCC), P.O. Box 12215, Research Triangle Park, NC 27709, <http://www.aatcc.org>.

<sup>5</sup> Available from U.S. Government Printing Office Superintendent of Documents, 732 N. Capitol St., NW, Mail Stop: SDE, Washington, DC 20401, <http://www.access.gpo.gov>.

#### 4. Summary of Test Method

4.1 This test method uses adult subjects who have provided a written informed consent and whose hands have been determined to be free from any apparent damage at the time of participation in the study. Subjects are to refrain from use of any antimicrobials for at least one week prior to the initiation of the test procedure (see Section 11).

4.2 Subjects' hands are artificially contaminated with 0.2 mL of a high-titer suspension of the test bacteria which is distributed over all surfaces of the hands and fingers to produce a minimum baseline recovery level of  $10^8$  cfu/hand. Because *Serratia marcescens* is relatively sensitive to drying, the high titer suspension is prepared by growing in broth with vigorous aeration, followed by a 10-fold concentration with centrifugation. *Staphylococcus aureus* is more resistant to drying and is therefore not concentrated after growth with vigorous aeration in broth.

4.3 Test material effectiveness is measured by comparing the number of test bacteria recovered from contaminated hands after use of the test material to the number recovered from contaminated hands not exposed to the test material. Activity of the test material is measured following a single application and after multiple consecutive contamination/application cycles in a single day. Evaluating effectiveness after multiple applications simulates repeated use of hand rubs in clinical settings and determines whether progressive build-up of non-volatile ingredients from the test material inhibits the antimicrobial action. An abbreviated test measuring activity of the test material following a single application may be used to simulate situations where high frequency use is not expected.

#### 5. Significance and Use

5.1 Hand hygiene is considered one of the most important measures for preventing the spread of infectious microorganisms. Hand rubs reduce the microbial load on the hands without the use of soap and water, and are thus an important tool in the practice of good hand hygiene. Alcohol-based hand rubs are recommended in healthcare settings for use on hands that are not visibly soiled. They are formulated to be applied full strength to dry hands, "rubbed in" until dry, and are not rinsed off.

5.2 This test method is designed specifically to evaluate hand rubs for efficacy in eliminating bacteria from experimentally-contaminated hands. It is designed as an alternative to Test Method E1174, which was intended primarily to evaluate antimicrobial handwashing agents that are lathered with the aid of water and then rinsed off. When using Test Method E1174 to evaluate hand rubs, inadequate drying of the hands after contamination dilutes the test material and can compromise activity, to result in an underestimation of effectiveness. Additionally, because hand rubs are not rinsed after product use, activity can be further degraded by build-up of soil from the contaminating broth and inactivated challenge bacteria on the hands.

5.2.1 In this method, application to the hands of a small volume of high-titer test bacteria suspension minimizes soil load such that the skin is completely dry prior to application of

the test material. Further, by applying the bacterial suspension only prior to those test material application cycles followed by sampling, excessive buildup of killed bacteria on the hands is avoided, and the potential impact of non-volatile test product ingredients on bacteria-eliminating effectiveness after ten consecutive applications can be specifically assessed.

5.3 A reference control is evaluated for each subject prior to evaluation of the test material. Data from the reference control helps to control for inter-subject variability, inter-experimental variability, and inter-laboratory variability; and enables improved statistical comparison of test materials evaluated in the same experiment.

5.4 This test method can be used to test any form of hand rubs, including gels, rinses, sprays, foams, and wipes when used according to label directions at typical "in-use" doses.

5.5 Susceptibility to biocides can vary among different species of bacteria and major differences have been noted between gram-negative and gram-positive organisms. This test method provides the option to use either a gram-negative bacterium (*Serratia marcescens*) or a gram-positive bacterium (*Staphylococcus aureus*) as the test organism. *S. marcescens* is used as a test organism in both Test Method E1174 and Test Method E2276. *S. aureus* is a highly relevant pathogen in healthcare, institutional, and community settings. Moreover, hands are an important vehicle in the transfer of *S. aureus* between people and the environment, and in the transfer between individuals.

5.6 This test method may be used as an alternative to Test Method E2276, which limits the test bacteria to the fingerpads and does not incorporate actual use conditions such as friction during hand rubbing.

5.7 The investigator should be aware of potential health risks associated with the use of these organisms and precautions similar to those referenced in Section 8 should be taken.

#### 6. Apparatus

6.1 *Centrifuge*—For the sedimentation of *S. marcescens* for concentration.

6.2 *Centrifuge Tubes*—Sterile, for sedimentation of *S. marcescens* for concentration.

6.3 *Colony Counter*—Any of several types may be used; for example, Quebec colony counters and similar devices. Automated, computerized plater/counter systems may also be used.

6.4 *Gloves*—Sterile, loose-fitting, unlined, powder-free gloves possessing no antimicrobial properties. Perform a zone of inhibition test, such as AATCC Test Method 147, to evaluate the antibacterial activity.

6.5 *Handwashing Sink*—Sufficient in size to permit handwashing without the touching of hands to sink surface or other subjects.

6.5.1 *Water Faucet(s)*—Located above the sink at a height to permit hands to be held higher than the elbow during the washing procedure.

6.5.2 *Tap Water Temperature Regulator and Temperature Monitor*—To set and maintain the tap water temperature at  $40 \pm 2^\circ\text{C}$ .

6.6 *Incubator*—Capable of maintaining temperatures of  $35 \pm 2^\circ\text{C}$  and  $25 \pm 2^\circ\text{C}$ . The latter temperature ensures adequate pigment production for *S. marcescens* on solid media.

6.7 *Miscellaneous Labware*—Continuously adjustable pipetters (1-mL and 0.2-mL capacity) and sterile pipette tips, sterile serological pipettes (5.0-mL capacity), sterile culture tubes, sterile disposable Petri dishes, sterile syringes, Erlenmeyer flasks, and beakers.

6.8 *Plastic Bags*—May be used in place of gloves (6.4). Bags should be approximately 29 by 31 cm, possess no antimicrobial properties, and have a low bioburden. Perform a zone of inhibition test, such as AATCC Test Method 147, to evaluate the antibacterial activity.

6.9 *Sampling Containers*—Sterile or sterilizable containers having tight closures and sufficient capacity to hold 75 mL sampling solution (see 7.7).

6.10 *Shaking Incubator*—Rotary platform shaking incubator capable of maintaining  $35 \pm 2^\circ\text{C}$  and capable of shaking at 250 r/min. Alternatively, use an incubator capable of maintaining  $35 \pm 2^\circ\text{C}$  and able to accommodate a portable rotary shaker, capable of shaking at 250 r/min.

6.11 *Sterilizer*—Any steam sterilizer capable of processing culture media and reagents.

6.12 *Timer (Stop-Clock)*—Type that can be read for minutes and seconds.

6.13 *Tourniquets*—Children's size or any style capable of securing gloves to the wrist.

6.14 *Vortex Mixer*—Any vortex that will ensure proper mixing of culture tubes.

## 7. Reagents and Materials

7.1 *Antibiotic Ointment*—A topical, triple-antibiotic ointment for application to the hands after the final decontamination.

7.2 *Cleansing Wash*—A mild, proven non-antimicrobial liquid soap. May be purchased commercially or prepared according to the instructions provided in Test Method E1174.

7.3 *Chlorhexidine Skin Cleanser*—Antiseptic skin cleanser containing 4 % chlorhexidine gluconate (w/v) for hand decontamination.

### 7.4 Culture Media:

7.4.1 *Broth*—Soybean-casein digest broth (tryptic soy broth) is recommended.

### 7.4.2 Agar Plating Media:

7.4.2.1 *S. aureus Plating Medium*—HardyCHROM (trademark), *Staph aureus*, available from Hardy Diagnostics, is recommended. Other indicator media for *S. aureus* or MRSA may be appropriate but should be validated prior to use.

NOTE 1—*S. aureus* forms smooth, deep pink to fuchsia-colored colonies. The growth of most other organisms, including *Staphylococcus epidermidis* are partially to completely inhibited.

7.4.2.2 *S. marcescens Plating Medium*—Soybean-casein digest agar (tryptic soy agar) is recommended.

7.5 *Dilution Fluid*—Sterile Butterfield's buffered phosphate diluent<sup>6</sup> (or other suitable diluent) adjusted to  $\text{pH } 7.2 \pm 0.1$  and containing an effective inactivator for the test material, if necessary.

NOTE 2—Inactivator is only required if neutralization of the test material cannot be achieved upon dilution into the sampling solution (see 7.7).

7.6 *Ethanol Solution*—70 % ethanol in water (v/v) for hand decontamination.

7.7 *Sampling Solution*—Dissolve 0.4 g  $\text{KH}_2\text{PO}_4$ , 10.1 g  $\text{Na}_2\text{HPO}_4$ , 1.0 g isooctylphenoxypolyethoxyethanol (for example, Triton X-100), and appropriately validated neutralizers, if necessary, in distilled water. Adjust pH to  $7.8 \pm 0.1$  with 0.1 N HCl or 0.1 N NaOH and bring volume to 1 L with distilled water. Sterilize in an autoclave and aseptically dispense 75-mL portions into sterile sampling containers (see 6.9).<sup>7</sup>

NOTE 3—A neutralizer validation should be conducted according to Test Methods prior to the study. Test Methods E1054 provides a list of neutralizers appropriate for commonly used antimicrobial agents. In some cases (for example, some alcohol-based hand rubs) neutralization is achieved by dilution alone.

7.8 *Test Material*—Use directions provided with the test material. If directions are not provided, use the directions given in this method.

7.9 *Reference Control*—60% isopropanol in water (v/v).

## 8. Test Bacteria

8.1 *Serratia marcescens* (ATCC 14756). This strain forms a stable red pigmentation at  $25^\circ\text{C}$ .

8.2 *Staphylococcus aureus* (ATCC 6538 (methicillin-sensitive) or ATCC 33591 (methicillin-resistant)) is an alternative test bacteria. *S. aureus* is differentiated from resident microbial skin flora (including *Staphylococcus epidermidis*) with chromogenic indicator medium (see 7.4.2.1). (**Warning**—Application of microorganisms to the skin may involve a health risk. Determine the antibiotic sensitivity profile of the test bacteria prior to applying to the skin. After the test has been completed, decontaminate the subject's hands and follow proper procedures to reduce infection risk (12.1 – 12.4). If an infection occurs, provide the antibiotic susceptibility profile to the attending clinician.)

## 9. Preparation of Test Bacteria Suspension

### 9.1 Method 1 (for *S. marcescens*):

9.1.1 A homogeneous bacterial suspension is used to inoculate the subjects' hands. Prepare a stock culture of *S. marcescens* (ATCC 14756) by inoculating approximately 5 mL of

<sup>6</sup> Horowitz, W., (Ed.), *Official Methods of Analysis of the AOAC International*, 18th Ed., Sec. 6.3.03 A.(f), Chapter 6, p. 10. AOAC International, Gaithersburg, MD, 2000.

<sup>7</sup> Peterson, A. F., "The Microbiology of the Hands: Evaluating the Effects of the Surgical Scrubs," *Developments in Industrial Microbiology*, Vol. 14, 1973, pp. 125–130.